

Terpenes from *Schisandra sphenanthera*

by Yan Jiang^a), Guang-Zhong Yang^{*a}), Yu Chen^b), Mao-Chuan Liao^a), Xiang-Ming Liu^a), Su Chen^a), Lu Liu^c), and Xin-Xiang Lei^{*c})

^a) Laboratory for Natural Product Chemistry, College of Pharmacy, South Central University for Nationalities, Wuhan 430074, P. R. China

(phone: +86-27-67841196; fax: +86-27-67841196; e-mail: yanggz888@126.com)

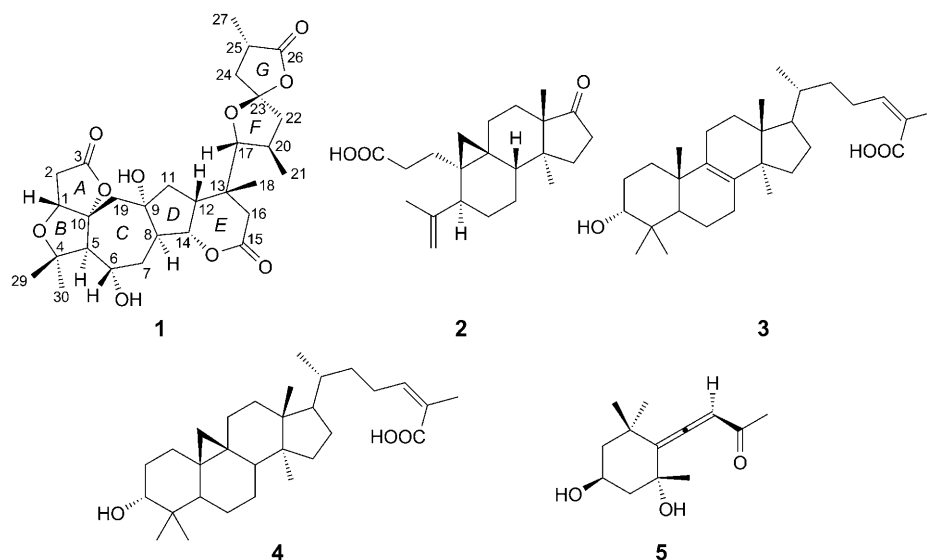
^b) College of Chemistry and Material Sciences, South Central University for Nationalities, Wuhan 430074, P. R. China

^c) College of Chemistry and Material Engineering, Wenzhou University, Wenzhou 325035, P. R. China
(phone: +86-0577-88373091; e-mail: xinxianglei@yahoo.com.cn)

One novel, highly oxygenated nortriterpenoid, schinrilactone C (**1**), and four known compounds, **2–5**, were isolated from the rattan of *Schisandra sphenanthera*. Their structures were determined by analysis of 1D- and 2D-NMR spectroscopic data. Schinrilactone C is the third example of wuweiziartane-type nortriterpenoids, bearing a modified five-membered *D* ring, a δ -lactone *E* ring, and a spirocyclic moiety in the side chain at C(13).

Introduction. – The economically and medicinally important family Schisandraceae, a family of climbing plants, contains the genera *Schisandra* and *Kadsura*. There are *ca.* 50 species in total in the world, and there are 29 species in China which are widely used as sedative and tonic agents in Traditional Chinese Medicine (TCM) [1]. The chemical constituents of Schisandraceae plants have been studied extensively. Previous phytochemical investigations on this family revealed lignans, especially dibenzocyclooctadienelignans with antihepatitis, antitumor, and anti-HIV activities [2–4]. In recent years, triterpenoids showing anti-HIV activities, and inhibitory activities toward cholesterol biosynthesis have also been isolated from this family [5].

Schisandra sphenanthera REHDER et E. H. WILSON is a climbing plant that is widely found in Sichuan, Hubei, Shanxi, and Yunnan Provinces. The fruits are used as antitussive and tonic agents under the name of *Nan-wuweizi* in TCM [6]. Phytochemical studies of *S. sphenanthera* fruits led to the isolation of six dibenzocyclooctadiene lignans, five other lignans, and three triterpenes, including schizandronic acid (ganwuweizic acid), anwuweizic acid, and schisanol [7]. Nine novel schisanartane-type nortriterpenoids, sphenadilactones A–C and sphenalactones A–D, a norcycloartane triterpenoid, sphenasin A, and a new phenolic glycoside have been isolated from the leaves and stems of *S. sphenanthera* [6][8]. In the course of our investigation on this genus, a novel highly oxygenated nortriterpenoid, schinrilactone C (**1**), together with four known compounds, micranoic acid B (**2**) [9], anwuweizic acid (**3**) [10], isoschizandronic acid (**4**) [11], and grasshopper ketone (**5**) [12] were isolated from the rattan of *S. sphenanthera* by normal-phase and reversed-phase silica-gel column chromatography. Their structures were identified by spectroscopic methods, especially 2D-NMR techniques.



Result and Discussion. – Schintrialactone C (**1**) was obtained as an amorphous white powder. Its empirical formula, $C_{29}H_{40}O_{10}$, was deduced, from HR-ESI-MS (m/z 571.2510 ($[M + Na]^+$; calc. 571.2519) and ^{13}C -NMR data, indicating ten degrees of unsaturation. Evident in the 1H -NMR spectrum were five Me signals due to three tertiary Me groups at $\delta(H)$ 0.92 (*s*), 1.28 (*s*), and 1.50 (*s*), and two secondary Me groups at $\delta(H)$ 0.71 (*d*, $J = 5.7$) and 0.92 (*d*, $J = 6.6$). Signals of four O-bearing CH groups appeared at $\delta(H)$ 3.41 (*d*, $J = 9.3$), 4.02–4.13 (*m*), 4.20 (*d*, $J = 4.5$), and 5.05 (*t*, $J = 8.7$). The ^{13}C -NMR and DEPT spectra of **1** exhibited signals for 29 C-atoms, including those of three ester groups ($\delta(C)$ 175.0, 169.5, and 178.6), five quaternary C-atoms (of three O-bearing C-atoms at $\delta(C)$ 85.7, 77.5, and 96.8 and one O–C–O group at $\delta(C)$ 112.1), nine CH groups (four O-bearing CH groups at $\delta(C)$ 82.1, 65.0, 83.5 and 90.6), seven CH_2 groups, and five Me groups. These features revealed that compound **1** was a highly oxygenated nortriterpene containing seven rings. Careful analysis of the 1H - and ^{13}C -NMR data (*Table*) revealed that **1** is similar to schintrialactone A, isolated before from *Schisandra chinensis* [13].

The structure of **1** was elucidated by analyzing the 2D-NMR data and by comparing these results with the NMR data reported for schintrialactone A. All these data revealed that **1** possesses a structure quite similar to that of schintrialactone A. However, different C- and H-atom chemical shifts were observed, indicating that the structures of **1** and schintrialactone A differ in ring C and the side chain at C(13). Specifically, the CH_2 group at C(6) in ring C of schintrialactone A was replaced by an O-bearing CH group in **1** with a downfield chemical shift at $\delta(C)$ 65.0. In the HMBC spectrum, correlations (*Table*) observed from Me(29) ($\delta(H)$ 1.28 (*s*)) to C(4) ($\delta(C)$ 85.7), C(5) ($\delta(C)$ 66.8), and C(30) ($\delta(C)$ 30.6), from Me(30) ($\delta(H)$ 1.50 (*s*)) to C(4) ($\delta(C)$ 85.7), C(5) ($\delta(C)$ 66.8), and C(29) ($\delta(C)$ 20.7), and from $CH_2(7)$ ($\delta(H)$ 2.37–2.43 (*m*)) to C(5) ($\delta(C)$ 66.8), C(6) ($\delta(C)$ 65.0), and C(8) ($\delta(C)$ 56.2), as well as the $^1H, ^1H$ -COSY spin system H–C(5)/H–C(6)/ $CH_2(7)$ /H–C(8), suggested that the OH group is located at C(6).

Table. ^1H - and ^{13}C -NMR, and HMBC Data (measured in $\text{C}_5\text{D}_5\text{N}$) of Compound **1**. δ in ppm, J in Hz.

	$\delta(\text{H})$	$\delta(\text{C})$	HMBC (H \rightarrow C)
H–C(1)	4.20 (<i>d</i> , $J = 4.5$)	82.1	C(2), C(3), C(10), C(19)
CH ₂ (2)	2.60 (overlapped, H _{α}), 2.74 (<i>d</i> , $J = 17.8$, H _{β})	35.3	
C(3)		175.0	
C(4)		85.7	
H–C(5)	2.30–2.50 (<i>m</i>)	66.8	
H–C(6)	4.02–4.13 (<i>m</i>)	65.0	
CH ₂ (7)	2.37–2.43 (<i>m</i>)	36.1	C(5), C(6), C(8), C(14)
H–C(8)	2.40–2.45 (<i>m</i>)	56.2	C(6), C(7), C(9), C(11)
C(9)		77.5	
C(10)		96.8	
CH ₂ (11)	2.14–2.20 (<i>m</i> , H _{α}), 1.98–2.06 (<i>m</i> , H _{β})	43.7	
H–C(12)	2.38–2.45 (<i>m</i>)	39.3	C(17)
C(13)		38.2	
H–C(14)	5.05 (<i>t</i> , $J = 8.7$)	83.5	C(15)
C(15)		169.5	
CH ₂ (16)	2.51 (<i>d</i> , $J = 14.4$, H _{α}), 2.27 (<i>d</i> , $J = 13.5$, H _{β})	35.6	H _{α} : C(15)
H–C(17)	3.41 (<i>d</i> , $J = 9.3$)	90.6	C(13), C(20), C(21), C(23), C(12)
Me(18)	0.92 (<i>s</i>)	14.9	C(12), C(13), C(16), C(17)
CH ₂ (19)	2.16–2.20 (<i>m</i>)	41.6	
H–C(20)	2.15–2.20 (<i>m</i>)	32.1	C(22), C(23)
Me(21)	0.71 (<i>d</i> , $J = 5.7$)	18.1	C(17), C(20), C(22)
CH ₂ (22)	2.04–2.10 (<i>m</i> , H _{α}), 1.40–1.45 (<i>m</i> , H _{β})	45.9	H _{α} : C(23), C(24)
C(23)		112.1	
CH ₂ (24)	1.73 (<i>t</i> -like, $J = 11.7$, H _{α}), 2.20–2.25 (<i>m</i> , H _{β})	40.3	C(22), C(23), C(25), C(26)
H–C(25)	2.64–2.71 (<i>m</i>)	35.1	C(24), C(26)
C(26)		178.6	
Me(27)	0.92 (<i>d</i> , $J = 6.6$)	18.7	C(24), C(25), C(26)
Me(29)	1.28 (<i>s</i>)	20.7	C(4), C(5)
Me(30)	1.50 (<i>s</i>)	30.6	C(4), C(5)

Further comparison of ^1H - and ^{13}C -NMR data with those of schinrilactone A, and analysis of 2D-NMR of **1** (Table) allowed us to identify rings A–E, leading to the establishment of partial structure **1a** (Fig. 1). A secondary Me resonance at $\delta(\text{H})$ 0.71 (*d*, $J = 5.7$) corresponding to Me(21) showed HMBC cross-peaks (Table) with H–C(20) ($\delta(\text{C})$ 32.1), and with H–C(17) ($\delta(\text{C})$ 90.6) and CH₂(22) ($\delta(\text{C})$ 45.9), which required that C(17) and C(22) are both attached to C(20) bearing the Me group. This was confirmed by the HMBCs between the O-bearing CH group at $\delta(\text{H})$ 3.41 (*d*, $J = 9.3$), corresponding to C(17), C(20), and C(21) ($\delta(\text{C})$ 18.1). Furthermore, ^1H , ^1H -COSY correlations, H–C(17)/H–C(20)/H–C(22) gave rise to partial structure **1b** (Fig. 1). HMBC Cross-peaks (Table) observed from Me(27) ($\delta(\text{H})$ 0.92 (*d*, $J = 6.6$)) to C(25) ($\delta(\text{C})$ 35.1), and to CH₂(24) ($\delta(\text{C})$ 40.3) and C(26) ($\delta(\text{C})$ 178.6), from H–C(25) ($\delta(\text{H})$ 2.64–2.71 (*m*)) to C(24) and C(26), and from H–C(24) ($\delta(\text{H})$ 2.20–2.25 (*m*)) to C(23) ($\delta(\text{C})$ 112.1), C(25), C(22) ($\delta(\text{C})$ 45.9), and C(26), along with the H-atom spin system deduced from ^1H , ^1H -COSY correlations H–C(24)/H–C(25)/Me(27) established the partial structure **1c** (Fig. 1).

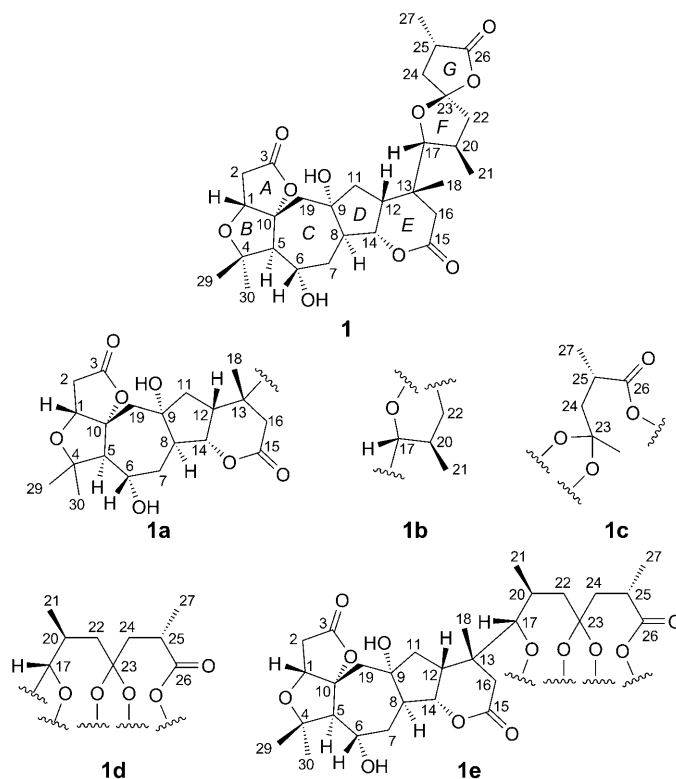


Fig. 1. The structure and structural fragments of **1**

Furthermore, HMBCs (Table) from CH₂(24) to C(22) and C(23), and from H–C(20) to C(22) and C(23) indicated direct connection of C(22) with C(23) and suggested fragment **1d** (Fig. 1). HMBCs (Table) from H–C(17) ($\delta(\text{H})$ 3.41 (*d*, $J = 9.3$)) to C(13) ($\delta(\text{C})$ 38.2) and from H–C(12) ($\delta(\text{H})$ 2.38–2.45 (*m*)) to C(17) ($\delta(\text{C})$ 90.6) required direct connection of C(13) with C(17) and permitted fragments **1a** and **1d** to be joined to provide **1e** (Fig. 1). On account of C(23), ($\delta(\text{C})$ 112.1) which was an O-bearing quaternary C-atom, connected with two O-atoms, at the two O-bridges between C(17)/C(23) and C(23)/C(26) were established.

The relative configuration of **1** was established from the ROESY spectrum (see Fig. 2). The relative configuration of rings A, B, and E of **1** was deduced to be the same as that in schintrilactone A from the similar C- and H-atom chemical shifts and ROESY correlations found in the spectra of both compounds. In the ROESY spectrum of **1**, correlations from Me(29) to H–C(1) and H–C(6), and from Me(30) to H–C(5) were observed, indicating that H–C(1) and H–C(6) are β -oriented, while H–C(5) and HO–C(6) possess α -orientation. In addition, the cross-peaks H–C(17)/Me(21), H–C(12), and H $_{\beta}$ –C(24), Me(18)/H–C(12), and Me(27)/H $_{\alpha}$ –C(24) in the ROESY spectrum demonstrated that H–C(12), Me(18), H–C(17), and Me(21) are β -oriented, while Me(27) possesses α -orientation. From these data, the structure and relative configuration of **1** was elucidated, and it was named as schintrilactone C (Fig. 2).

Fr. 3 (20 g) was subjected to CC (SiO₂; cyclohexane/CHCl₃/MeOH 9:1:0, 8:2:0, 7:3:0, 1:1:0, 0:1:0, 0:98:2): Frs. 3.1–3.4. Fr. 3.3 (4.4 g) was subjected to CC (ODS; H₂O/MeOH 7:3→0:1): Frs. 3.3.1–3.3.7. *Micranoic acid B* (**2**; 17.3 mg) was crystallized from Fr. 3.3.5. Fr. 5 (13.7 g) was subjected to CC (SiO₂; cyclohexane/CHCl₃/acetone 2:8:0, 1:9:0, 0:1:0, 0:95:5, 0:9:1, 0:8:2, 0:6:4): Frs. 5.1–Fr. 5.9. Fr. 5.6 (5.0 g) was subjected to CC (ODS, H₂O/MeOH 7:3→0:1): Frs. 5.6.1–5.6.9. Fr. 5.6.2 (439 mg) was subjected to CC (SiO₂; cyclohexane/CHCl₃/MeOH 3:7:0, 2:8:0, 1:9:0, 0:1:0, 0:98:2, 0:95:5, 0:9:1, 0:8:2, 0:7:3, 0:0:1): Frs. 5.6.2.1–5.6.2.9. Fr. 5.6.2.3 was purified by semi-prep. HPLC (MeOH/H₂O 30:70, 3ml/min) to give *grasshopper ketone* (**5**; 8.9 mg) at *t*_R 25 min. Fr. 5.6.5 (369 mg) was subjected to CC (SiO₂; cyclohexane/CHCl₃/MeOH 3:7:0, 2:8:0, 1:9:0, 0:1:0, 0:98:2, 0:95:5, 0:9:1, 0:8:2, 0:7:3, 0:0:1): *schinrilactone C* (**1**; 11.6 mg).

Schinrilactone C (=rel-(3*a*R,5*a*S,6*S*,7*a*R,7*b*S,11*R*,11*a*R,12*a*S,13*a*R)-11-[(2*R*,3*R*,5*R*,8*S*)-3,8-*Di*-methyl-7-oxo-1,6-dioxaspiro[4.4]non-2-yl]-tetradecahydro-6,12*a*-dihydroxy-5,5,11-trimethyl-2*H*,9*H*-furo[3',2':2,3]furo[3,4':5,6]azuleno[1,2-*b*]pyran-2,9-dione; **1**). Amorphous white powder. [*a*]_D = +55 (*c* = 0.125, pyridine). ¹H- and ¹³C-NMR: see the *Table*. HR-ESI-MS: 571.2510 (C₂₉H₄₀NaO₇; calc. 571.2519).

This work was financially supported by *Academic Team of South Central University for Nationalities: Ethno-Medicine Research and Development* (XTZ09010).

REFERENCES

- [1] W.-L. Xiao, R.-T. Li, S.-X. Huang, J.-X. Pu, H.-D. Sun, *Nat. Prod. Rep.* **2008**, *25*, 871.
- [2] H.-Y. Min, E.-J. Park, J.-Y. Hong, Y.-J. Kang, S.-J. Kim, H.-J. Chung, E. R. Woo, T. M. Hung, U. F. Houn, U. J. Youn, Y. S. Kim, S. S. Kang, K. H. Bae, S. K. Lee, *Bioorg. Med. Chem.* **2008**, *18*, 523.
- [3] D.-F. Chen, S.-X. Zhang, M. Kozuka, Q.-Z. Sun, J. Feng, Q. Wang, T. Mukainaka, Y. Nobukuni, H. Tokuda, H. Nishino, H. K. Wang, S. L. Morris-Natschke, K.-H. Lee, *J. Nat. Prod.* **2002**, *65*, 1242.
- [4] X.-M. Gao, J.-X. Pu, S.-X. Huang, L.-M. Yang, H. Huang, W.-L. Xiao, Y.-T. Zheng, H.-D. Sun, *J. Nat. Prod.* **2008**, *71*, 558.
- [5] W.-L. Xiao, R.-R. Tian, J.-X. Pu, X. Li, L. Wu, Y. Lu, S.-H. Li, R.-T. Li, Y.-T. Zheng, Q.-T. Zheng, H.-D. Sun, *J. Nat. Prod.* **2006**, *69*, 277.
- [6] R. T. Li, Z. Y. Weng, J. X. Pu, H. D. Sun, *Chin. Chem. Lett.* **2008**, *19*, 696.
- [7] Y. Lu, D.-F. Chen, *J. Chromatogr. A* **2009**, *1216*, 1980.
- [8] W.-L. Xiao, S.-X. Huang, R.-R. Wang, J.-L. Zhong, X.-M. Gao, F. He, J.-X. Pu, Y. Lu, Y.-T. Zheng, Q.-T. Zheng, H.-D. Sun, *Phytochemistry* **2008**, *69*, 2862.
- [9] R.-T. Li, Q.-B. Han, A.-H. Zhao, H.-D. Sun, *Chem. Pharm. Bull.* **2003**, *51*, 1174.
- [10] J.-S. Liu, M.-F. Huang, *Acta. Chim. Sin.* **1984**, *42*, 264.
- [11] J. S. Liu, W. G. Wang, *Chin. Tradit. Herbal Drugs* **1990**, *21*, 294.
- [12] T. Miyase, A. Ueno, N. Takizawa, H. Kobayashi, H. Karasawa, *Chem. Pharm. Bull.* **1987**, *35*, 1109.
- [13] S.-X. Huang, J. Yang, H. Huang, L.-M. Li, W.-L. Xiao, R.-T. Li, H.-D. Sun, *Org. Lett.* **2007**, *9*, 4175.
- [14] C. Lei, S.-X. Huang, J.-J. Chen, L.-B. Yang, W.-L. Xiao, Y. Chang, Y. Lu, H. Huang, J.-X. Pu, H.-D. Sun, *J. Nat. Prod.* **2008**, *71*, 1228.
- [15] C. Lei, J.-X. Pu, S.-X. Huang, J.-J. Chen, J.-P. Liu, L.-B. Yang, Y.-B. Ma, W.-L. Xiao, X.-N. Li, H.-D. Sun, *Tetrahedron* **2009**, *65*, 164.
- [16] C. Lei, S.-X. Huang, J.-J. Chen, J.-X. Pu, L.-M. Li, W.-L. Xiao, J.-P. Liu, L.-B. Yang, H.-D. Sun, *Helv. Chim. Acta* **2007**, *90*, 1399.
- [17] S.-X. Huang, R.-T. Li, J.-P. Liu, Y. Lu, Y. Chang, C. Lei, W.-L. Xiao, L.-B. Yang, Q.-T. Zheng, H.-D. Sun, *Org. Lett.* **2007**, *9*, 2079.
- [18] J.-X. Pu, R.-T. Li, W.-L. Xiao, N.-B. Gong, S.-X. Huang, Y. Lu, Q.-T. Zheng, L.-G. Lou, H.-D. Sun, *Tetrahedron* **2006**, *62*, 6073.

Received June 27, 2010